

PROCHAMAZULENE SESQUITERPENE LACTONES FROM *STEVIA SERRATA*

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Key Word Index—*Stevia serrata*; Compositae; Eupatorieae; sesquiterpene lactones; guaianolides; prochamazulenes.

Abstract—The leaves of *Stevia serrata* afforded two new sesquiterpene lactones. Their structures and stereochemistry were established by spectral methods, mainly ^1H NMR.

INTRODUCTION

In the course of the column chromatographic separation of the components from *Stevia serrata* Cav. on silica gel, we noticed the presence of chamazulene in the non-polar fractions. This observation led to the isolation of two new prochamazulene sesquiterpene lactones, **1a** and **1b**, and the known lactones christinine I and III [1, 2]. To our knowledge, the only three known prochamazulenes are artabsin, matricin and 4-epimatricin from *Artemisia arborescens* [3] and *A. absinthium* [4].

RESULTS AND DISCUSSION

Steviserrolide A (**1a**), $\text{C}_{17}\text{H}_{22}\text{O}_5$, was an unstable colourless oil, which exhibited the typical IR absorption band of a γ -lactone at 1773 cm^{-1} . Further absorptions at 3696, 3577 and 1736 cm^{-1} indicated the presence of hydroxyl and ester functions, respectively.

The ^1H NMR spectrum (Table 1) of **1a** lacked the typical doublets of the exocyclic methylene conjugated with the γ -lactone, which must be saturated as indicated by a doublet at $\delta 1.20$ ($J = 7.0\text{ Hz}$). The two doublets at $\delta 6.45$ and 5.90 ($J = 5.5\text{ Hz}$) were assigned to the double bond at C-2 in the five-membered ring of the guaiane skeleton.

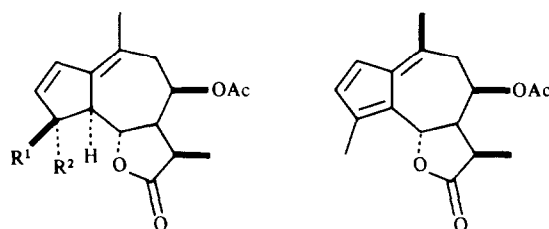
A sharp singlet at $\delta 2.03$ in the ^1H NMR spectrum, together with strong mass spectral peaks at m/z 43 and $246 [\text{M} - \text{HOAc}]^+$ indicated the presence of an acetate group. Their position and stereochemistry could be deduced from the small coupling constants and the downfield chemical shift observed for H-8 ($\delta 5.28$), which were almost identical to those of 11,13-dihydroguaianolides with a β -oriented acetate group [1, 5] and markedly different to those of 11,13-dihydroguaianolides with an α -oriented ester group [3, 6]. Assuming the α -orientation of H-7, as in all the sesquiterpene lactones of higher plants, [7], the stereochemistry at C-5 and C-6 was assigned based on the coupling constants of the corresponding protons ($J_{5,6} = 10\text{ Hz}$, $J_{6,7} = 10\text{ Hz}$) confirming the *trans* ring attachment of the γ -lactone. The assignment of the configuration of the C-11 methyl group in **1a** as β pseudoaxial was suggested on the basis of the observed

Table 1. ^1H NMR spectral data for compounds **1a**, **1b** and **2** (80 MHz, CDCl_3 , TMS as int. standard)

H	1a	1b	2
2	6.45 <i>d</i> (6.08)	6.32 <i>d</i>	6.50 <i>d</i>
3	5.90 <i>d</i> (5.78)	5.90 <i>d</i>	6.20 <i>d</i>
6	4.82 <i>t</i> (4.50)	4.70 <i>t</i>	5.72 <i>br d</i>
8	5.28 <i>br d</i> (4.90)	5.27 <i>m</i>	5.40 <i>t</i>
13	1.20 <i>d</i> (0.90)	1.20 <i>d</i>	1.20 <i>d</i>
14	1.78 <i>br s</i> (1.50)	1.78 <i>br s</i>	2.15 <i>br s</i>
15	1.57 <i>s</i> (1.40)	1.48 <i>s</i>	2.15 <i>br s</i>
OAc	2.03 <i>s</i> (1.55)	2.05 <i>s</i>	2.05 <i>s</i>

J (Hz) **1a**: 2,3 = 5.5; 5,6 = 6,7 = 10; 7,8 = 8,9 ~ 1.5; 8,9' = 5.0; 11,13 = 7.0; **1b**: 8,9' = 3.0; **2**: 2,3 = 6.0; 6,7 = 10; 7,8 = 0; 8,9 = 8,9' = 4.0; 11,13 = 7.0.

Numbers in parentheses are chemical shifts in C_6D_6 .



1a $\text{R}^1 = \text{OH}$, $\text{R}^2 = \text{Me}$

1b $\text{R}^1 = \text{Me}$, $\text{R}^2 = \text{OH}$

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solvent shift [8], ($\Delta\delta\text{ CDCl}_3 - \text{C}_6\text{D}_6$) = 0.30. This value was identical to those of christinine I [1] and deacetoxy-matricin [6], and larger than those published for matricin ($\Delta\delta = 0.20$) and 4-epimatricin ($\Delta\delta = 0.18$) [3], both with an α pseudo equatorial methyl group at C-11. The configuration at C-4 was established from the observed chemical shifts for H-6 and H-15 in similar C-4 hydroxy epimeric guaianolides [3, 9]. In agreement with the β -orientation

of the C-4 hydroxyl group in **1a**, the chemical shifts for H-6 and H-15 were shifted to lower field than those of **1b**.

Steviserrolide B (**1b**), $C_{17}H_{22}O_5$ is also a colourless oil. Comparison of the 1H NMR spectra of **1b** and **1a** (Table 1), clearly showed differences for the proton signals at C-2, C-15 and C-6, indicating that both are epimers at C-4. Consequently, the structure for steviserrolide B is depicted as in **1b**.

Addition of trichloroacetyl isocyanate to the lactone **1b** in the NMR sample tube, gave the fulvene **2**. This elimination instead of acylation of the hydroxyl group at C-4, has been also observed for matricin [3, 11] under the same conditions. Finally, an inspection of a Dreiding model of **2**, clearly showed that the dihedral angle value between H-7 and H-8 was nearly 90° , in agreement with the $J_{7,8}$ value of zero.

EXPERIMENTAL

Stevia serrata Cav., was collected near Chapala, State of Jalisco, in September 1983, and near Cuernavaca, State of Morelos, on the Mexico-Cuernavaca road (Km 64), in August 1983. Voucher specimens were deposited in the Herbarium of the Instituto de Biología, UNAM.

Isolation of christinines and chamazulene. Dried leaves (187 g) of *S. serrata* from Chapala, were extracted with petrol at room temp. The petrol extract (8.7 g) was separated by CC over silica gel (270 g) using petrol and petrol-EtOAc as eluants, to yield 39 fractions. Fraction 7 (85 mg) was purified by prep. TLC (petrol) affording 47 mg of chamazulene [10]. Fractions 29–31 eluted with petrol-EtOAc (1:1) were combined to give 1.18 g. A 450 mg sample was purified by prep. TLC (CH_2Cl_2 -Me₂CO 19:1) yielding 126 mg of christinine III [2] and 45 mg of christinine I [1].

Chamazulene. 1H NMR (80 MHz, $CDCl_3$): δ 1.31 (3H, J = 7.0 Hz, $-CH_2-Me$), 2.63 (3H, s, C-1-Me), 2.80 (3H, s, C-4-Me), 2.83 (2H, q, J = 7.0 Hz, $-CH_2-Me$), 6.9 (1H, d, J = 11 Hz, H-5), 7.17 (1H, d, J = 4 Hz, H-3), 7.35 (1H, dd, J = 11, 1.5 Hz, H-6), 7.68 (1H, d, J = 4 Hz, H-2), 8.12 (1H, d, J = 1.5 Hz, H-8); EIMS (probe) 70 eV m/z (rel. int.): 184 $[M]^+$ (100), 169 $[M-Me]^+$ (86), 155 (30), 153 (35).

Isolation of steviserrolides A and B. Due to the extreme sensitivity of the pro-chamazulene lactones to weak acid conditions [4], these were separated using prep. TLC plates, with loss of large amounts of material. Dried leaves (155 g) of *S. serrata*

from Cuernavaca were extracted with CH_2Cl_2 at room temp. and the extract was evapd *in vacuo* to give a greenish syrup (9.75 g). A 0.75 g sample was separated by prep. TLC (CH_2Cl_2 -Me₂CO, 19:1, $\times 2$) to give 75 mg of crude **1a** and 67 mg of crude **1b**. Further purification using analytical TLC plates (CH_2Cl_2 -Me₂CO, 9:1, $\times 2$) afforded 19 mg of **1a** and 15 mg of **1b**.

Steviserrolide A (1a). Unstable colourless oil. IR $\nu_{max}^{CHCl_3}$ cm^{-1} : 3696, 3577, 1773, 1736, 1601; EIMS 70 eV, m/z (rel. int.): 306 $[M]^+$ (2.3), 246 $[M-HOAc]^+$ (33), 231 $[M-HOAc-Me]^+$ (100), 185 (24), 173 (36), 43 $[Me-CO]^+$ (78).

Steviserrolide B (1b). Unstable colourless oil. IR $\nu_{max}^{CHCl_3}$ cm^{-1} : 3686, 3592, 1774, 1737, 1602; EIMS 70 eV m/z (rel. int.): 306 $[M]^+$ (1.6), 246 $[M-HOAc]^+$ (18), 231 $[M-HOAc-Me]^+$ (26), 145 (23), 43 $[MeCO]^+$ (100).

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REFERENCES

1. Salmón, M., Díaz, E. and Ortega, A. (1973) *J. Org. Chem.* **38**, 1759.
2. Salmón, M., Díaz, E. and Ortega, A. (1977) *Rev. Latinoam. Quím.* **8**, 172.
3. Appendino, G. and Gariboldi, P. (1982) *Phytochemistry* **21**, 2555.
4. Vokac, K., Herout, V. and Sorm, F. (1969) *Coll. Czech. Chem. Commun.* **34**, 2288.
5. Bohlmann, F. and Zdero, C. (1985) *Liebigs Ann. Chem.* 1764.
6. Bohlmann, F., Borthakur, N., Jakupovic, J. and Pickard, J. (1982) *Phytochemistry* **21**, 1357.
7. Fischer, N. H., Olivier, E. J. and Fischer, H. D. (1979) *Progress in the Chemistry of Organic Natural Products* (Herz, W., Grisebach, H. and Kirby, G. B., eds), Vol. 38, p. 47. Springer, Vienna.
8. Narayanan, C. R. and Venkatasubramanian, N. K. (1968) *J. Org. Chem.* **33**, 3156.
9. Bohlmann, F. and Knoll, K. H. (1979) *Phytochemistry* **18**, 995.
10. Bertelli, D. J. and Crabtree, J. H. (1968) *Tetrahedron* **24**, 2079.
11. Samek, Z. and Budesinsky, M. (1979) *Coll. Czech. Chem. Commun.* **44**, 558.